

Synthesis, Characterization, and DNA-Binding Studies of Nitro(oligopyridine)ruthenium(II) Complexes

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The complexes of general formulas $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-CO}_2\text{H-4}'\text{-Mebpy})(\text{X})]^{n+}$ ($\text{X} = \text{NO}$ ($n = 3$) and NO_2 ($n = 1$); **1**, **2**) and $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-COGHK-4}'\text{-Mebpy})(\text{X})]$ ($\text{X} = \text{NO}$ ($n = 3$) and NO_2 ($n = 1$); **3**, **4**) were synthesized and characterized. The complex $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-CO}_2\text{-4}'\text{-Mebpy})(\text{NO}_2)] \cdot 7.5\text{H}_2\text{O}$ has also been characterized by X-ray crystallographic studies. It crystallizes in the triclinic system: $a = 9.4982(1) \text{ \AA}$, $b = 13.1330(1) \text{ \AA}$, $c = 14.2498(2) \text{ \AA}$; $\alpha = 110.5870(6)^\circ$, $\beta = 98.4048(5)^\circ$, $\gamma = 106.4353(5)^\circ$, $P\bar{1}$, $Z = 2$. The crystal structure reveals an extended hydrogen-bonding network. Two water molecules form strong hydrogen bonds with the nitro and the carboxylic oxygen atoms of two separate units of the complex, resulting in a dimeric unit. The dimers are bridged by a $(\text{H}_2\text{O})_{15}$ cluster, consisting of two *cyclo*-(H_2O)₆ species, while an *exo*- $\text{H}_2\text{O}(8)$ connects them. Two more *exo*- H_2O molecules are joined together and connect the *cyclo*-(H_2O)₆ units with the $\text{H}_2\text{O}(1)$ of the dimeric unit. It was found that complexes **1** and **3** can be transformed into their nitro derivatives in aqueous media at neutral pH. Photorelease of NO in dry MeCN solutions was observed for complexes **1** and **3**. Also, complex **2** partially releases $(\text{NO}_2)^-$ in MeCN upon visible light irradiation. Complex **2** interacts with short fragments (70–300 bp) of calf thymus DNA shortening slightly the apparent polynucleotide length, while the conjugation of the peptide GHK to it (**2**) affects its DNA-binding mode. The peptide moiety of complex **4** was found to interact with the DNA helix in a synergistic way with the whole complex. Preliminary results of photocleavage of DNA by complex **2** are also reported.

Introduction

A large number of ruthenium nitrosyl complexes containing polypyridine ligands, such as 2,2'-bipyridine (bpy) or 2,2':6',2''-terpyridine (terpy), have been synthesized and extensively studied over the past few years.^{1–4} Among them,

complexes with possible uses as regulators of blood pressure^{5,6} or as antitumor agents releasing the NO within tumor cells^{7,8} have been suggested. However, a crucial question, concerning to the stability of the coordinated NO in aqueous media of biological systems, still remains. An exciting question is whether such complexes exist as nitrosyl or whether they are transformed into their nitro forms. It seems that it depends not only on the nature of the ruthenium coordination sphere, but also on the nature of the co-ligands.

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Thus, the nitrosyl complexes $[\text{Ru}^{\text{II}}(\text{trpy})(\text{dpk})\text{NO}]^{3+}$ (dpk = 2,2'-dipyridylketone),⁹ $[\text{Ru}^{\text{II}}(\text{trpy})(\text{L})\text{NO}]^{3+}$ (L = 2,2'-dipyridylamine),¹⁰ and $[\text{Ru}^{\text{II}}(\text{trpy})(\text{L}')\text{NO}]^{3+}$ (L' = various substituted azopyridine ligands)¹¹ are stable in the solid state and in dry organic solvents, but in contact with water they are transformed into their nitro derivatives. On the other hand, similar complexes such as $[\text{Ru}^{\text{II}}(\text{trpy})(\text{L}')\text{NO}]^{3+}$ (L' = 2-(2-pyridyl)azole derivatives) are unusually stable in aqueous media.¹² In fact, all the above complexes have the same ruthenium coordination sphere consisting of five nitrogen donor atoms from pyridine rings and the nitrogen atom from NO. Moreover, ruthenium nitrosyls with a similar coordination sphere may photorelease the coordinated NO in aqueous media.^{13,14}

The noncoordination DNA binding and the photoreactivity of ruthenium polypyridine complexes have been the subject of research interest in the past few years.^{15–19} These coordination compounds are able to bind to DNA in different modes (electrostatically, surface or groove binding, intercalation) depending on the ligand size and nature, and the chirality of the octahedral complex. The most studied compound, $[\text{Ru}(\text{bpy})_3]^{2+}$, shows a rather weak association with DNA, most probably due to its electrostatic interactions with the DNA phosphates.^{20–22} In order to enhance the DNA binding of $[\text{Ru}(\text{bpy})_3]^{2+}$, peptides or amino acids have been tethered to a bpy ligand, forming chimerical complexes, able to bind with higher affinity to DNA, and assisted by an additional peptide–DNA interaction.^{23–30} The nature of the peptide affects both the DNA binding affinity and the cross-

linking efficiency of the ruthenium–peptide conjugated complex.^{28–30} Concerning the DNA photocleavage, it has been suggested that the distance between the bound complex and the DNA helix plays a key role, since the life of the active oxygen species generated by the photoactivation of the ruthenium polypyridine complex is strongly distance dependent.^{31,32} For this reason, complexes containing ligands that bind tightly to DNA (e.g., intercalators) have shown an acceleration of the photocleavage process compared to groove bound complexes.³¹ However, in the case of the chimeric complex $[\text{Ru}(\text{bpy})_2(\text{phenIA})](\text{PF}_6)_2$ (phenIA = *N*-iodoacetyl-5-amino-1,10-phenanthroline) containing a 54-residue peptide, which is conjugated to the phenIA ligand, no photo-induced DNA damage was observed.³³

In the present report an attempt to investigate how polypyridine nitrosyl ruthenium complexes interact with the DNA upon photoactivation is described. For this reason, the complexes $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-CO}_2\text{H-4'-Meppy})(\text{X})]^{n+}$ (X = NO ($n = 3$) and NO₂ ($n = 1$); **1**, **2**) and $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-COGHK-4'-Meppy})(\text{X})]$ (X = NO ($n = 3$) and NO₂ ($n = 1$); **3**, **4**) were synthesized and characterized by spectroscopic and crystallographic techniques. The photochemistry of complexes **1**, **2**, and **3** was studied in dry MeCN, as was their stability in aqueous media. Since only the nitro derivatives were found to be stable, DNA binding interactions were studied with complexes **2** and **4** in order to investigate the role of the conjugated peptide in the DNA-binding mode. Finally, complex **2** was irradiated with visible light to examine its DNA photocleavage.

Experimental Section

(i). Materials and Methods. 2,2':6',2''-Terpyridine was purchased from Aldrich Chemical Co. and used without further purification. The complex $\text{Ru}(\text{terpy})\text{Cl}_3$ ³⁴ and the ligand 4-carboxy-4'-methyl-2,2'-bipyridine (4-CO₂H-4'-Meppy)²⁷ were prepared according to literature procedures. The complex $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-CO}_2\text{H-4'-Meppy})\text{Cl}](\text{PF}_6)$ was prepared and purified by a literature method.³⁵ The complex $[\text{Ru}(\text{terpy})(4\text{-COGHKCONH}_2\text{-4'-Meppy})\text{Cl}](\text{PF}_6)$ was prepared by solid-phase synthesis as previously described.³⁶ The sonication of the calf thymus DNA was performed according to the literature.³⁷ The DNA concentration, expressed as moles of nucleotides per liter [P], was determined on a Pharmacia LKB-Biochrom 4060 UV–vis spectrophotometer from the absorbance at 260 nm ($\epsilon_{60} = 6600 \text{ M}^{-1} \times \text{cm}^{-1}$, $T = 298 \text{ K}$). The sample was found to be consistent with low protein content as the value of the A_{260}/A_{280} ratio was 1.9.

The infrared spectra of the complexes were recorded on a Perkin-Elmer Paragon 1000 FTIR spectrophotometer, equipped with a

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Golden Gate Diamond ATR device, using the diffuse reflectance technique. C, H, and N determinations were performed on a Perkin-Elmer 2400 series II analyzer. The ^{23}Na NMR and ^{31}P NMR spectra were obtained on a Bruker 300 DPX spectrometer operating at 121.49 and 79.39 MHz for ^{31}P and ^{23}Na NMR, respectively, at 310 K in D_2O solutions. The LC-MS experiments were performed on a Finnigan MAT TSQ-700 instrument with a custom-made electrospray interface (ESI) combined with LC-equipment consisting of a Dionex pump P580, a Gilson 119 UV-detector at 214 and 254 nm, and an Alltima C18 150 mm \times 4.6 mm i.d. column. UV-vis spectra were recorded on a Varian Cary 3-Bio spectrophotometer with temperature controller. The irradiation of the solutions was carried out with a Philips mercury lamp. CD spectra were recorded on a Jobin Yvon CD-6 instrument at room temperature.

(ii). Synthesis of the Complexes. Synthesis of $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-CO}_2\text{H-4'-Mebpy})\text{NO}](\text{PF}_6)_3$ (1). A 500 mg (0.68 mmol) portion of $[\text{Ru}(\text{terpy})(4\text{-CO}_2\text{H-4'-Mebpy})\text{Cl}](\text{PF}_6)$ was suspended in 10 mL of water, and the solution was refluxed for 15 min. The deep red-brown solution was filtered hot and then cooled to room temperature. The pH of the solution was adjusted to ~ 2 with the appropriate amount of 1 M HCl. A 1 mL portion of aqueous solution containing 47 mg (0.68 mmol) of sodium nitrite, NaNO_2 , was added slowly over a 10 min period. After complete addition, the brown-red solution became yellow and was warmed at 60 $^\circ\text{C}$ for an additional 45 min. A 4 mL solution of saturated NH_4PF_6 was added, and the precipitate was collected by filtration, washed with diethyl ether, and dried in vacuo (yield 58%). $\text{C}_{27}\text{H}_{21}\text{F}_{18}\text{N}_6\text{O}_3\text{P}_3\text{Ru}$ (%) calcd: C 32.00, H 2.09, N 8.29. Found: C 31.34, H 2.23, N 8.23.

Synthesis of $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-CO}_2\text{H-4'-Mebpy})\text{NO}_2](\text{PF}_6)_3$ (2). A 500 mg (0.68 mmol) portion of $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-CO}_2\text{H-4'-Mebpy})\text{Cl}](\text{PF}_6)$ was suspended in 10 mL of water, and the solution was refluxed for 15 min. The deep red-brown solution was filtered hot and then cooled to room temperature. The pH of the solution was adjusted to ~ 10 with 1M NaOH, and then 1 mL of an aqueous solution containing 47 mg (0.68 mmol) of sodium nitrite, NaNO_2 , was added dropwise over a 10 min period. The brown-red solution becomes orange, and it is warmed at 60 $^\circ\text{C}$ for an additional 30 min. After cooling at room temperature, a 4 mL solution of saturated NH_4PF_6 was added. The precipitate was collected by filtration, washed with diethyl ether, and dried in vacuo (yield 52%). ESI-MS: m/z 595, $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-CO}_2\text{H-4'-Mebpy})(\text{NO}_2)]^+$. $\text{C}_{27}\text{H}_{21}\text{F}_6\text{N}_6\text{O}_4\text{PRu}$ (%) calcd: C 43.85, H 2.86, N 11.36. Found: C 43.34, H 2.63, N 11.02. ^1H NMR (300 MHz, D_2O) isomer I: $\delta = 2.62$ (s, 3H, bpy-Me), 9.45 (d, 1H, bpy-H6), 8.08 (d, 1H, bpy-H5), 8.75 (s, 1H, bpy-H3), 7.32 (d, 1H, bpy-H6'), 7.21 (d, 1H, bpy-H5'), 8.43 (s, 1H, bpy-H3'), 8.27 (d, 2H, tpy-H6H6'), 7.84 (t, 2H, tpy-H5H5'), 7.18 (t, 1H, tpy-H4), 7.18 (t, 1H, tpy-H4'), 7.62 (d, 1H, tpy-H3), 7.62 (d, 1H, tpy-H3'), 8.41 (d, 1H, tpy-H3''), 8.17 (m, 1H, tpy-H4''), 8.41 (d, 1H, tpy-H5'').

Isomer II: $\delta = 2.24$ (s, 3H, bpy-Me), 7.07 (d, 1H, bpy-H6), 6.79 (d, 1H, bpy-H5), 8.14 (s, 1H, bpy-H3), 9.17 (d, 1H, bpy-H6'), 7.68 (d, 1H, bpy-H5'), 8.51 (s, 1H, bpy-H3'), 8.27 (d, 2H, tpy-H6H6'), 7.84 (t, 2H, tpy-H5H5'), 7.18 (t, 1H, tpy-H4), 7.18 (t, 1H, tpy-H4'), 7.62 (d, 1H, tpy-H3), 7.62 (d, 1H, tpy-H3'), 8.42 (d, 1H, tpy-H3''), 8.17 (m, 1H, tpy-H4''), 8.42 (d, 1H, tpy-H5'').

Synthesis of $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-COGHKCONH}_2\text{-4'-Mebpy})(\text{NO})](\text{PF}_6)_3$ (3). The synthesis of complex 3 was made in a similar way to complex 1. A 100 mg (0.106 mmol) portion of $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-COGHKCONH}_2\text{-4'-Mebpy})\text{Cl}]\text{Cl}$ and 0.5 mL of an aqueous solution containing 7.3 mg of sodium nitrite, NaNO_2 , (0.106 mmol) were used for this reaction (yield 55%). $\text{C}_{41}\text{H}_{44}\text{F}_{18}\text{N}_{13}\text{O}_3\text{P}_3\text{Ru}$ (%) calcd C, 36.89; H, 3.32; N, 13.64. Found: C, 36.97; H, 3.43; N, 13.12. FTIR (KBr): $\nu(\text{NO}) = 1924\text{ cm}^{-1}$ (s).

Synthesis of $[\text{Ru}^{\text{II}}(4\text{-COGHKCONH}_2\text{-4'-Mebpy})(\text{NO}_2)](\text{PF}_6)_3$ (4). The synthesis of complex 4 was performed in a similar fashion to complex 2. A 150 mg (0.16 mmol) portion of $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-COGHKCONH}_2\text{-4'-Mebpy})\text{Cl}]\text{Cl}$ and 1 mL of aqueous solution containing 11.04 mg (0.16 mmol) of sodium nitrite, NaNO_2 , were used for this reaction (yield 60%). ESI-MS: m/z 916 $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-COGHKCONH}_2\text{-4'-Mebpy})\text{NO}_2]^+$. $\text{C}_{41}\text{H}_{44}\text{F}_6\text{N}_{13}\text{O}_6\text{PRu}$ (%) calcd C, 46.42; H, 4.18; N, 17.16. Found: C, 46.17; H, 4.13; N, 17.09. ^1H NMR (300 MHz, D_2O) isomer I: $\delta = 3.02$ (s, 3H, bpy-Me), 9.29 (d, 1H, bpy-H6), 7.79 (d, 1H, bpy-H5), 8.32 (s, 1H, bpy-H3), 8.51 (d, 1H, bpy-H6'), 8.25 (d, 1H, bpy-H5'), 8.21 (s, 1H, bpy-H3'), 8.10 (d, 2H, tpy-H6H6'), 7.65 (t, 2H, tpy-H5H5'), 8.07 (t, 1H, tpy-H4), 8.07 (t, 1H, tpy-H4'), 8.31 (d, 1H, tpy-H3), 8.31 (d, 1H, tpy-H3'), 7.27 (d, 1H, tpy-H3''), 7.73 (m, 1H, tpy-H4''), 7.27 (d, 1H, tpy-H5''), 8.35 (s, 1H, H2-His), 6.78 (s, 1H, H5-His), 4.70 (d, 1H, α -His), 3.58 (t, 1H, β -His), 3.94 (s, 1H, α -Gly), 3.50 (d, 1H, α -Lys), 1.97 (m, 1H, β -Lys), 1.47 (m, 1H, γ -Lys), 2.04 (m, 1H, δ -Lys), 3.02 (t, 1H, ϵ -Lys).

Isomer II: $\delta = 2.73$ (s, 3H, bpy-Me), 8.37 (d, 1H, bpy-H6), 7.93 (d, 1H, bpy-H5), 7.29 (s, 1H, bpy-H3), 9.04 (d, 1H, bpy-H6'), 7.65 (d, 1H, bpy-H5'), 8.05 (s, 1H, bpy-H3'), 7.96 (d, 2H, tpy-H6H6'), 7.27 (t, 2H, tpy-H5H5'), 7.93 (t, 1H, tpy-H4), 7.93 (t, 1H, tpy-H4'), 8.35 (d, 1H, tpy-H3), 8.35 (d, 1H, tpy-H3'), 7.23 (d, 1H, tpy-H3''), 7.71 (m, 1H, tpy-H4''), 7.23 (d, 1H, tpy-H5''), 8.32 (s, 1H, H2-His), 6.75 (s, 1H, H5-His), 4.66 (d, 1H, α -His), 3.53 (t, 1H, β -His), 3.84 (s, 1H, α -Gly), 3.17 (d, 1H, α -Lys), 1.60 (m, 1H, β -Lys), 1.40 (m, 1H, γ -Lys), 2.00 (m, 1H, δ -Lys), 2.96 (m, 1H, ϵ -Lys).

(iii) Generation of Photoproducts for FTIR Analysis. The 2 mg portions of the complexes $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-CO}_2\text{H-4'-Mebpy})\text{NO}](\text{PF}_6)_3$ (1) and $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-COGHKCONH}_2\text{-4'-Mebpy})\text{NO}](\text{PF}_6)_3$ (3) were dissolved in degassed MeCN (0.5 mL) and irradiated with light from a mercury lamp for 2 h. A stream of argon for the evaporation of the solvent was used, leaving a solid orange residue that was analyzed by infrared spectroscopy.

(iv) Generation of Photoproducts for UV-Vis Analysis. Solutions (10^{-4} M) in degassed MeCN of the complexes $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-CO}_2\text{H-4'-Mebpy})\text{NO}](\text{PF}_6)_3$, $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-CO}_2\text{H-4'-Mebpy})\text{NO}_2](\text{PF}_6)_3$, and $[\text{Ru}^{\text{II}}(\text{terpy})(4\text{-COGHKCONH}_2\text{-4'-Mebpy})\text{NO}](\text{PF}_6)_3$ were irradiated with a mercury lamp for 3 h. The complexes were dissolved in degassed MeCN (10^{-4} mol L^{-1}), and the solutions were placed in a 1 cm path length quartz cuvette. The photochemistry of $[\text{Ru}(\text{terpy})(4\text{-CO}_2\text{H-4'-Mebpy})\text{NO}](\text{PF}_6)_3$ was also studied in phosphoric buffer at pH = 3. Successive spectra were recorded to monitor the photolysis of the complexes.

X-ray Crystal Structure Determination of 2. X-ray intensities were measured on a Nonius Kappa CCD diffractometer with rotating anode (graphite monochromator, $\lambda = 0.71073\text{ \AA}$) up to a resolution of $(\sin \theta/\lambda)_{\text{max}} = 0.65\text{ \AA}^{-1}$ at a temperature of 150 K. The structure was solved with Direct Methods³⁸ and refined with SHELXL-97³⁹ against F^2 of all reflections. Non-hydrogen atoms were refined with anisotropic displacement parameters. All hydrogen atoms were located in the difference Fourier map. C-H hydrogen atoms were refined with a riding model; O-H hydrogen atoms were kept fixed on their located positions. The methyl group at C17 was found rotationally disordered. Water molecule O8 was refined with occupancy of 0.5. Geometry calculations and checking

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Table 1. Experimental Details of the Crystal Structure Determination of **2**

formula	C ₂₇ H ₂₀ N ₆ O ₄ Ru·7.5 H ₂ O
fw	728.68
cryst color	red
cryst size [mm ³]	0.12 × 0.12 × 0.09
cryst syst	triclinic
space group	P $\bar{1}$ (No. 2)
a [Å]	9.4982(1)
b [Å]	13.1330(1)
c [Å]	14.2498(2)
α [deg]	110.5870(6)
β [deg]	98.4048(5)
γ [deg]	106.4353(5)
V [Å ³]	1535.12(3)
Z	2
d_{calc} [g/cm ³]	1.576
μ [mm ⁻¹]	0.582
abs correction	SORTAV ^[63]
abs correction range	0.90–0.95
reflns (measured/unique)	24813/6942
R_{int}	0.0475
params/restraints	416/0
R1/wR2 [$I > 2\sigma(I)$]	0.0341/0.0838
R1/wR2 [all reflns]	0.0447/0.0899
S	1.065
$\rho_{\text{min/max}}$ [e/Å ³]	–0.62/1.02

for higher symmetry were performed with the PLATON program.⁴⁰ Further experimental details, like absorption correction,⁴¹ are given in Table 1.

Preparation of Ruthenium Adducts with DNA for Electronic and CD Spectra. The interactions of complexes [Ru^{II}(terpy)(4-CO₂H-4'-Mebpy)NO₂](PF₆) and [Ru^{II}(terpy)(4-COGHKCONH₂-4'-Mebpy)NO₂](PF₆) with calf thymus DNA were studied with UV–vis and CD spectroscopy by adding the required volume of a freshly prepared solution of the complexes dissolved in 1 mM PIPES and 20 mM sodium chloride aqueous solution, to calf thymus DNA solutions (100 μ M) and incubating at 25 °C for 24 h. The final ruthenium/DNA base pair ratios (r) were 0.1, 0.3, and 0.5 (complex concentration was 10, 30, and 50 μ M). CD spectra were also recorded for the study of the irradiation of the system [Ru^{II}(terpy)(4-CO₂H-4'-Mebpy)NO₂](PF₆)–DNA at ratio $r = 0.1$. Samples of DNA with complex **2** were prepared as above. CD spectra were recorded before and after the irradiation of the solutions with the mercury lamp for 30 min. In this experiment, the effect of the irradiated complex to DNA was also studied. In that case, the sample of complex [Ru^{II}(terpy)(4-CO₂H-4'-Mebpy)NO₂](PF₆) was irradiated with the mercury lamp and subsequently was mixed with the DNA sample.

Results and Discussion

(i) Synthesis and Characterization of the Complexes.

Complexes **1–4** were synthesized following the replacement of the chloride in the corresponding chlorido complexes and the adjustment of the pH of the reaction to the desired value. The nitrosyl complexes were obtained in acidic conditions (pH \sim 2) and the nitro in basic conditions (pH \sim 10). The IR spectra of the prepared nitrosyl complexes **1** and **3** show strong and sharp IR bands at 1955 and 1924 cm⁻¹, respectively, which were assigned to the stretching vibration frequency $\nu(\text{NO})$. The $\nu(\text{NO})$ stretching frequency of **1** is very similar to that observed in the complex [Ru^{II}(terpy)-

Table 2. Diagnostic Spectroscopic Data for Complexes **1–4**

no.	electronic spectral data λ_{max} (nm)	
	in dry MeCN.	infrared spectral data (cm ⁻¹)
1	250 $\pi \rightarrow \pi^*$ (intraligand)	$\nu(\text{NO}) = 1955$
	289 $d_{\pi}(\text{Ru}) \rightarrow \pi^*(\text{terpy})$	$\nu(\text{CO}) = 1706$
	304 $d_{\pi}(\text{Ru}) \rightarrow \pi^*(\text{bpy})$	
	339 $d_{\pi}(\text{Ru}) \rightarrow \pi^*(\text{NO})$	
2	269, 280 $\pi \rightarrow \pi^*$ (intraligand),	$\nu(\text{CO}) = 1732$
	304 $d_{\pi}(\text{Ru}) \rightarrow \pi^*(\text{terpy})$	$\nu_{\text{as}}(\text{NO}_2) = 1419$
	329 $d_{\pi}(\text{Ru}) \rightarrow \pi^*(\text{bpy})$,	$\nu_{\text{s}}(\text{NO}_2) = 1272$
	475 $d_{\pi}(\text{Ru}) \rightarrow \pi^*(\text{NO}_2)$	
3	270 $\pi \rightarrow \pi^*$ (intraligand)	$\nu(\text{NO}) = 1924$
	310 $\pi \rightarrow \pi^*$ (intraligand)	$\nu(\text{CO}) = 1645$
	314 $d_{\pi}(\text{Ru}) \rightarrow \pi^*(\text{terpy})$	
	325 $d_{\pi}(\text{Ru}) \rightarrow \pi^*(\text{bpy})$	
4	350 $d_{\pi}(\text{Ru}) \rightarrow \pi^*(\text{NO})$	
	280 $\pi \rightarrow \pi^*$ (intraligand),	$\nu(\text{CO}) = 1648$
	306 $d_{\pi}(\text{Ru}) \rightarrow \pi^*(\text{terpy})$	$\nu_{\text{as}}(\text{NO}_2) = 1437$
	330 $d_{\pi}(\text{Ru}) \rightarrow \pi^*(\text{bpy})$,	$\nu_{\text{s}}(\text{NO}_2) = 1263$
	473 $d_{\pi}(\text{Ru}) \rightarrow \pi^*(\text{NO}_2)$	

(bpy)NO³⁺ (1952 cm⁻¹).⁴² Also, a strong band at 1706 cm⁻¹ assigned to the $\nu(\text{C}=\text{O})$ stretching vibration indicates that the carboxyl group of the ligand 4-CO₂H-4'-Mebpy is protonated. The conjugation of the peptide GHK in complex **3** affects the value of the $\nu(\text{NO})$ stretching frequency, which is shifted to lower frequencies by about 30 cm⁻¹. In general, the value of $\nu(\text{NO})$ is a good indicator of the degree of positive charge residue on the coordinated NO.⁴³ Thus, this value decreases upon replacement of the –CO₂H with –COGHK at the para position of the pyridine ring in the ligand 4-CO₂H-4'-Mebpy, indicating that the electrophilic reactivity of the nitrosyl group could be decreased from complex **3** to **1**. This does not seem to be expected, since the electrophilic reactivity of the ruthenium nitrosyl complexes depends on the nature of the inner-sphere coordinated ligands rather than the substituents on the pyridine ring.⁴⁴ In the IR spectra of nitro complexes **2** and **4**, the absence of the $\nu(\text{NO})$ stretching vibration band indicates the complete transformation of the corresponding nitrosyl complexes to the nitro ones. Also, the $\nu(\text{C}=\text{O})$ band appeared at 1660 and 1676 cm⁻¹ in the spectra of complexes **2** and **4**, respectively, indicating that the carboxyl group of the ligand 4-CO₂H-4'-Mebpy in both complexes is deprotonated.

In the electronic spectra of complexes **1** and **3** in dry MeCN solutions, the intense bands at 250 and 274 nm correspondingly are most likely intraligand $\pi \rightarrow \pi^*$ transitions.^{42,45} The bands at 289 and 304 nm in the spectrum of complex **1**, which coincide in a broad band at 310 in the case of complex **3**, are probably MLCT transitions $d_{\pi}\text{Ru} \rightarrow \pi^*(\text{ligand})$.^{42,46} The lowest energy bands at 339 and 350 nm in the spectra of **1** and **3**, correspondingly, are attributed to the $d_{\pi}\text{Ru} \rightarrow \pi^*(\text{NO})$ transition.^{42,46} The band shifting to lower energies from **1** to **3** reflects the destabilization of the d_{π} level, due to the $d_{\pi}\text{Ru} \rightarrow \pi^*(\text{NO})$ back-bonding, and thus, the electrophilic reactivity decreases going from **3** to **1**. On

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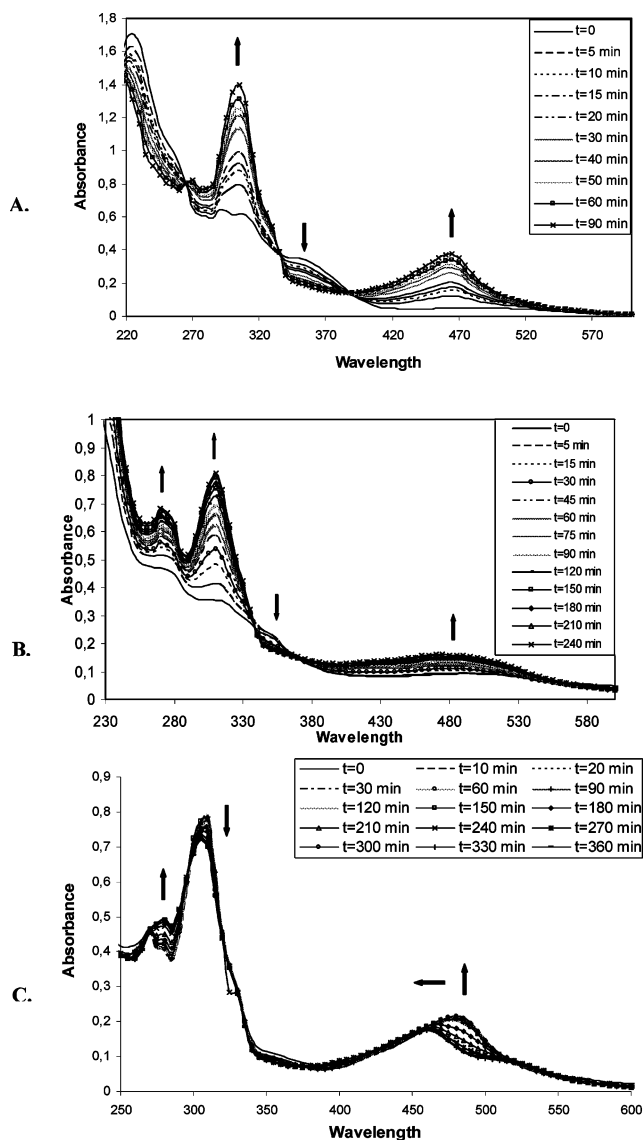


Figure 1. Electronic spectra of complexes **1**, **3**, and **2** (A, B, and C, respectively), in dry degassed 10^{-4} M MeCN solutions during light irradiation. Arrows indicate increasing or decreasing of the absorbance intensity.

the other hand, the nitro complexes **2** and **4** display the MLCT band $d_{\pi}\text{Ru} \rightarrow \pi^*(\text{NO}_2)$ red-shifted at 475 and 473 nm, correspondingly. All relevant spectral data are summarized in Table 2.

The purity of complexes **2** and **4** was checked by mass spectrometry (MALDI TOF and ESI MS) using MeCN as a carrier solvent. However, in the cases of nitrosyl complexes **1** and **3**, the moisture traces present in the carrier solvent partially transformed them to the corresponding nitro complexes.¹¹

(ii) Photochemistry of the Complexes. The photochemistry of nitrosyl complexes **1** and **3** was studied in carefully dried MeCN solution (10^{-4} M) using light irradiation with a mercury lamp (150 W). Upon the irradiation, the light orange color of the solutions turned to brown accompanied by changes in their UV–vis spectra. Successive scanning was taken in 10 min intervals, and the original bands of the complexes were replaced by new ones (Figure 1). Figure

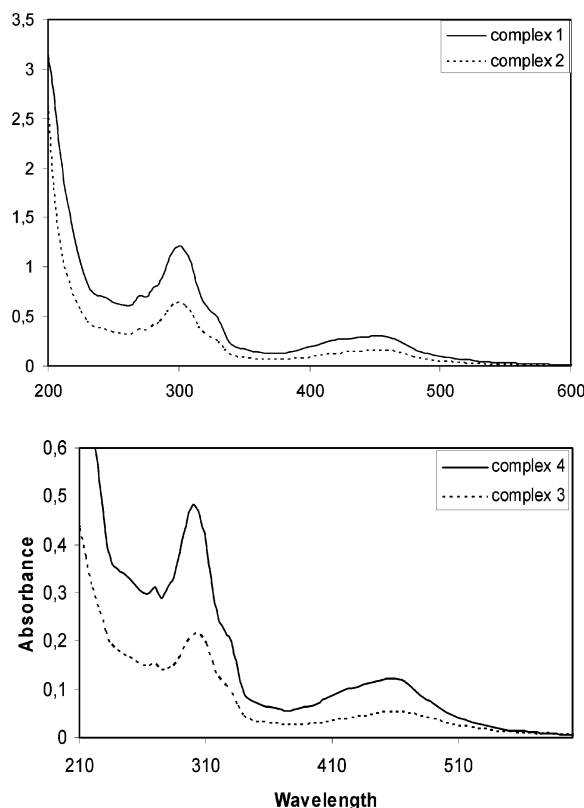


Figure 2. (A) Electronic spectra of complexes **1** and **2** under the same pH conditions in consistent buffer at 1 mM PIPES and 20 mM sodium chloride at 25 °C. (B) Electronic spectra of complexes **3** and **4** under the same pH conditions in consistent buffer at 1 mM PIPES and 20 mM sodium chloride at 25 °C.

1A demonstrates the spectral changes observed for complex **1**. These changes display a clear transformation of the complex to a photoproduct, characterized by three isosbestic points at 265, 334, and 390 nm indicating that possible intermediates or byproducts are not in significant concentrations to make measurable changes. The MLCT absorption band at 339 nm ($d_{\pi}\text{Ru} \rightarrow \pi^*(\text{NO})$) decreases, indicative of NO photorelease (Figure 1A). A new low-energy absorption band at 464 nm is appearing which arises from a $d_{\pi}\text{Ru} \rightarrow \pi^*(\text{MeCN})$ and/or from a $\pi(\text{ligand}) \rightarrow d_{\pi}\text{Ru}$ transition.⁴⁷ The bands at 289 and 304 nm ($d_{\pi}\text{Ru} \rightarrow \pi^*(\text{ligand})$) were observed broadened and clustered together at 302 nm, while their intensity increased dramatically. The intraligand $\pi \rightarrow \pi^*$ transition bands were red-shifted to 269 nm (see Table 1) which is characteristic of the formation of Ru(III) species.⁴⁵ In addition, the EPR spectrum of the photoproduct showed paramagnetic species, suggesting the presence of Ru(III). Exhaustive irradiation of the solution leads to a final stable spectrum after 2 h. Similar spectral changes were observed upon irradiation of the MeCN solution of complex **3** (Figure 1B). Two isosbestic points at 338 and 372 nm were observed, indicating the transformation of the complex to a photoproduct. The MLCT absorption band at 350 nm increases, while a new broad band at 473 nm is appearing. The intensities of the intraligand $\pi \rightarrow \pi^*$ and the MLCT $d_{\pi}\text{Ru} \rightarrow \pi^*(\text{ligand})$ bands increase, indicating that the replacement

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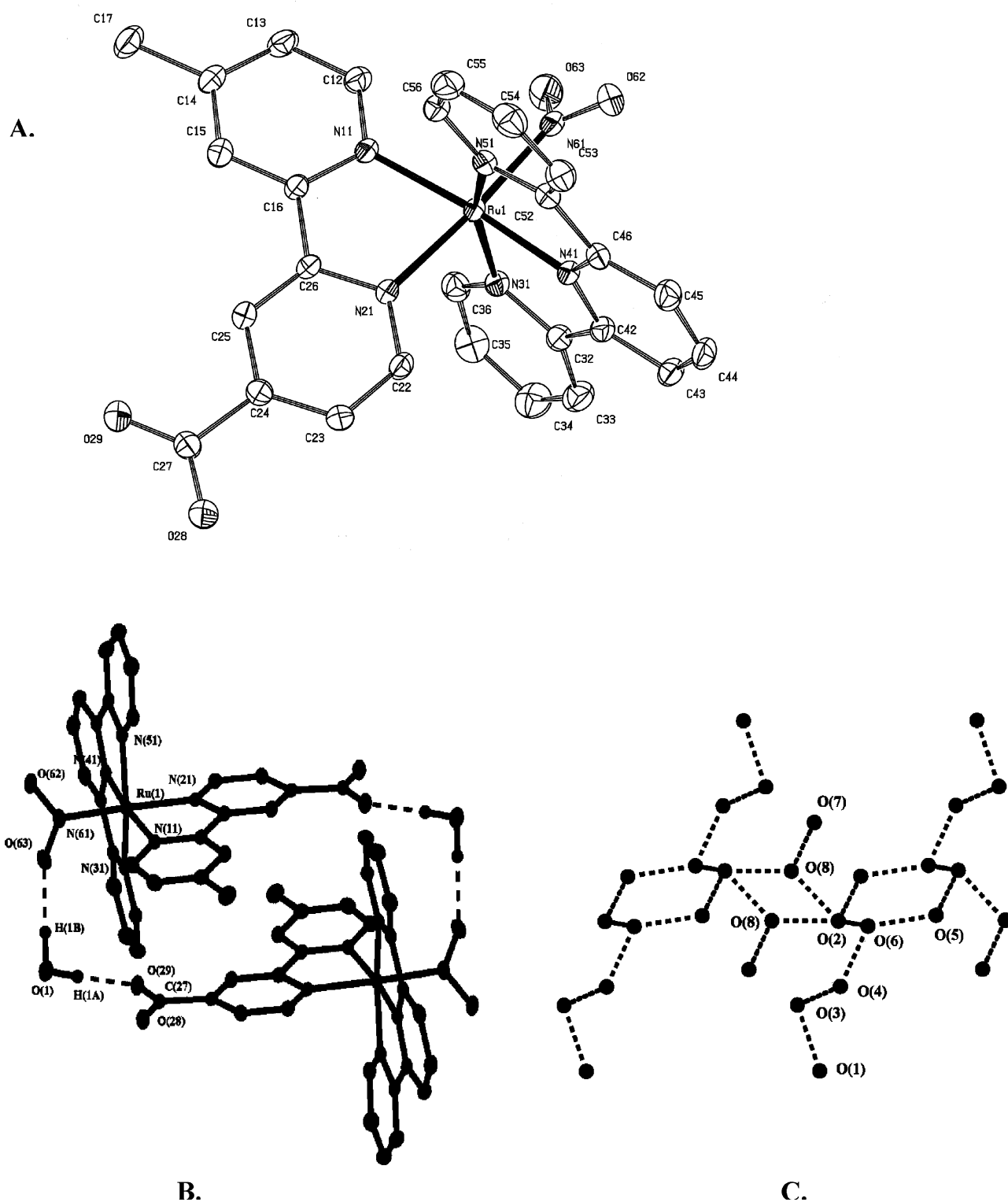


Figure 3. (A) ORTEP diagram of deprotonated complex **2** with 50% probability thermal ellipsoids showing the atomic labeling scheme. (B) Hydrogen bonding of the dimer. (C) Supramolecular $(\text{H}_2\text{O})_{15}$ network.

of the NO is followed by oxidation of the ruthenium center. The NO dissociation from the precursor complex *cis*-(Cl,
Cl)[Ru^{II}Cl₂(NO⁺)(terpy)]Cl has been previously reported, suggesting a weak coordination of the MeCN in addition to the oxidation of the ruthenium center.³⁶

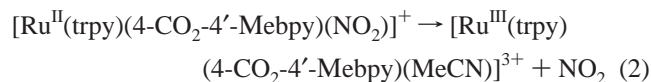
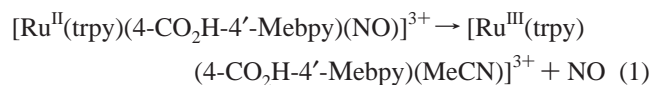
The photochemistry of the nitro complex (**2**) was studied in MeCN solution during the irradiation with a mercury lamp

until a final stable spectrum (6 h) was obtained (Figure 1C). Spectral changes in the UV and visible region were observed since the complex was transformed to a photoproduct, characterized by two isosbestic points at 324 and 515 nm. The $d_{\pi} \rightarrow \pi(\text{NO}_2)$ absorption band is decreasing, and two new bands are appearing at 505 and 464 nm. The MLCT band at 464 nm is very similar to that observed in the case

Table 3. Selected Bond Distances [Å] and Angles [deg] for the Deprotonated Complex **2**

N11–Ru1	2.0961(19)	N11–Ru1–N51	98.46(8)
N21–Ru1	2.0708(19)	N11–Ru1–N61	97.11(8)
N31–Ru1	2.078(2)	N21–Ru1–N31	89.77(8)
N41–Ru1	1.9669(19)	N21–Ru1–N41	96.02(8)
N51–Ru1	2.065(2)	N21–Ru1–N51	90.63(8)
N61–Ru1	2.029(2)	N21–Ru1–N61	175.34(7)
N61–O62	1.261(3)	N31–Ru1–N41	79.48(8)
N61–O63	1.250(3)	N31–Ru1–N51	159.31(8)
O62–N61–O63	116.7(2)	N31–Ru1–N61	91.42(8)
N11–Ru1–N21	78.23(7)	N41–Ru1–N51	79.90(8)
N11–Ru1–N31	101.87(8)	N41–Ru1–N61	88.63(8)
N11–Ru1–N41	174.04(8)	N51–Ru1–N61	89.85(8)

of complex **1** and could be attributed to the $d_{\pi}\text{Ru} \rightarrow \pi^*$ -(MeCN) transition, while the lowest energy band at 505 nm is assigned as the MLCT band $d_{\pi} \rightarrow \pi$ (ligand). In the EPR spectrum of the final photoproduct, paramagnetic species corresponding to Ru(III) were observed. LC-MS spectrometric analysis of the irradiated solution is showing the formation of a solvolyzed product. The singly charged positive ion at m/z 675.7 corresponding to the $\text{C}_{31}\text{H}_{26}\text{N}_8\text{O}_4\text{-Ru}$ could be assigned to the solvolyzed cation $\{[\text{Ru}^{\text{III}}(\text{trpy})\text{-}(4\text{-CO}_2\text{-4}'\text{-Mebpy})(\text{MeCN})]\text{NO}_2\cdot\text{MeCN}\}^+$, while the related protonated double charged cation $\{[\text{Ru}^{\text{III}}(\text{trpy})(4\text{-CO}_2\text{H-4}'\text{-Mebpy})(\text{MeCN})]\text{NO}_2\cdot\text{MeCN}\}^{2+}$ appears at m/z 338.4. Also, a signal at m/z 379.4 corresponding to the $\text{C}_{29}\text{H}_{23}\text{N}_6\text{O}_2\text{F}_6\text{-PNaRu}$ could be assigned to the doubly charged sodium-containing cation $\{[\text{Ru}^{\text{III}}(\text{trpy})(4\text{-CO}_2\text{Na-4}'\text{-Mebpy})(\text{MeCN})]\text{(PF}_6\text{)}\}^{2+}$. The presence of the Na^+ is likely originating from the addition of NaOH during the transformation of the precursor chlorido-complex to the nitro derivative. Conclusively in all cases studied, the results confirmed the photooxidation of Ru(II) to Ru(III) and the dissociation of the NO and NO_2 in MeCN solutions according to the following reactions:



(iii) Aqueous Media Studies of the Complexes. The frequencies of the $\nu(\text{NO})$ stretching vibration in complexes **1** and **3** (1955 and 1924 cm^{-1}) indicate that the degree of the positive charge residue on the coordinated NO is high.⁴³ Such high values reflect the electrophilic reactivity of the NO group. The nucleophilic attack of the hydroxyl group to the NO converts the nitrosyl complexes to corresponding nitro, through eq 3.



In order to study the DNA interactions of complexes **1–4** in aqueous media, the stability of the above compounds at neutral pH was investigated. As presented in Figure 2, the electronic spectra of complexes **1** and **2** (Figure 2A) are exactly the same, and this is also true for complexes **3** and

Table 4. Hydrogen Bonding Interactions in the Crystal Structure of **2**

D–H···A	D–H	H···A	D···A	D–H···A
O1–H1A···O29 ^{ia}	0.93	1.80	2.727(3)	172
O1–H1B···O63 ⁱⁱ	1.02	1.94	2.957(3)	173
O2–H2A···O28	0.91	1.84	2.743(3)	172
O2–H2B···O5 ⁱⁱⁱ	0.99	1.88	2.794(3)	152
O3–H3A···O28 ⁱ	0.97	1.75	2.716(3)	173
O3–H3B···O1	0.92	1.99	2.869(3)	160
O4–H4A···O62 ^{iv}	0.95	1.94	2.890(3)	172
O4–H4B···O3 ^v	0.93	1.86	2.784(3)	172
O5–H5A···O2 ⁱⁱⁱ	1.02	1.80	2.794(3)	163
O5–H5B···O6	0.95	1.93	2.829(4)	159
O6–H6A···O4 ^{vi}	0.95	1.83	2.736(4)	158
O6–H6B···O2	1.06	1.90	2.868(4)	151
O7–H7A···O8 ^{vi}	0.96	2.01	2.973(8)	174
O7–H7B···O62 ^{vii}	1.01	2.02	3.021(3)	168
O8–H8A···O2 ^v	1.00	1.85	2.810(8)	162
O8–H8B···O2 ⁱⁱⁱ	1.02	2.08	2.802(9)	126

^a Water molecule O8 was refined with an occupancy of 0.5. Symmetry operations follow: i, 2 – x, 1 – y, 1 – z; ii, 1 + x, y, z; iii, 1 – x, –y, 1 – z; iv, –x, 1 – y, –z; v, x – 1, y, z; vi, 1 – x, 1 – y, 1 – z; vii, 1 – x, 1 – y, –z.

4 (Figure 2B). This effect clearly shows that nitrosyl complexes **1** and **3** were transformed to the corresponding nitro complex at neutral pH.

The ¹H NMR spectra of complexes **1** and **2** in D₂O (buffer phosphates, pH = 7.0) are the same. They show double resonances for the protons of ligand 4-CO₂-4'-Mebpy and slightly broadened signals for the terpy, indicating the presence of two chemically nonequivalent species. Assuming that only the nitro complex is present at this pH, a possible explanation could be the existence of two positional isomers (differing in the orientation of the ligand 4-CO₂H-4'-Mebpy, toward ruthenium center).³⁵ Double resonances for all protons of the ligand 4-CO₂H-4'-Mebpy in the spectra of complexes **3** and **4** were also observed. The ¹H NMR signals of complexes **1–4** were assigned, assisted by TOCSY experiments.

(iv) Crystal and Molecular Structure of Deprotonated Complex 2. The molecular structure of **2** is shown in Figure 3A. Table 3 presents selected bond distances and angles. The crystal structure of the complex consists of neutral units of the complex and water molecules involved in a network of hydrogen bonds. Two symmetry related water molecules O1 and O1' link two complex molecules via the nitro oxygen atoms O(63)/O(63)' and O(29)/O(29)' to form a centrosymmetric, hydrogen-bonded dimer. The hydrogen bonds in this dimer are strong with O···O distances of 2.717(3) and 2.957(3) Å, respectively (Table 4, Figure 3B). The dimers are bridged to other dimers by (H₂O)₁₅ clusters, involving two *cyclo*-(H₂O)₆ connected by an *exo*-H₂O(8) in a half occupancy. Two more *exo*-H₂O species are joined together connecting the *cyclo*-(H₂O)₆ units with the H₂O(1) of the dimer (Figure 3C). Similar *cyclo*-(H₂O)₆ in a chairlike conformation have been observed recently in the crystal lattice of various metal complexes.^{48,9,49,50} In the cyclic

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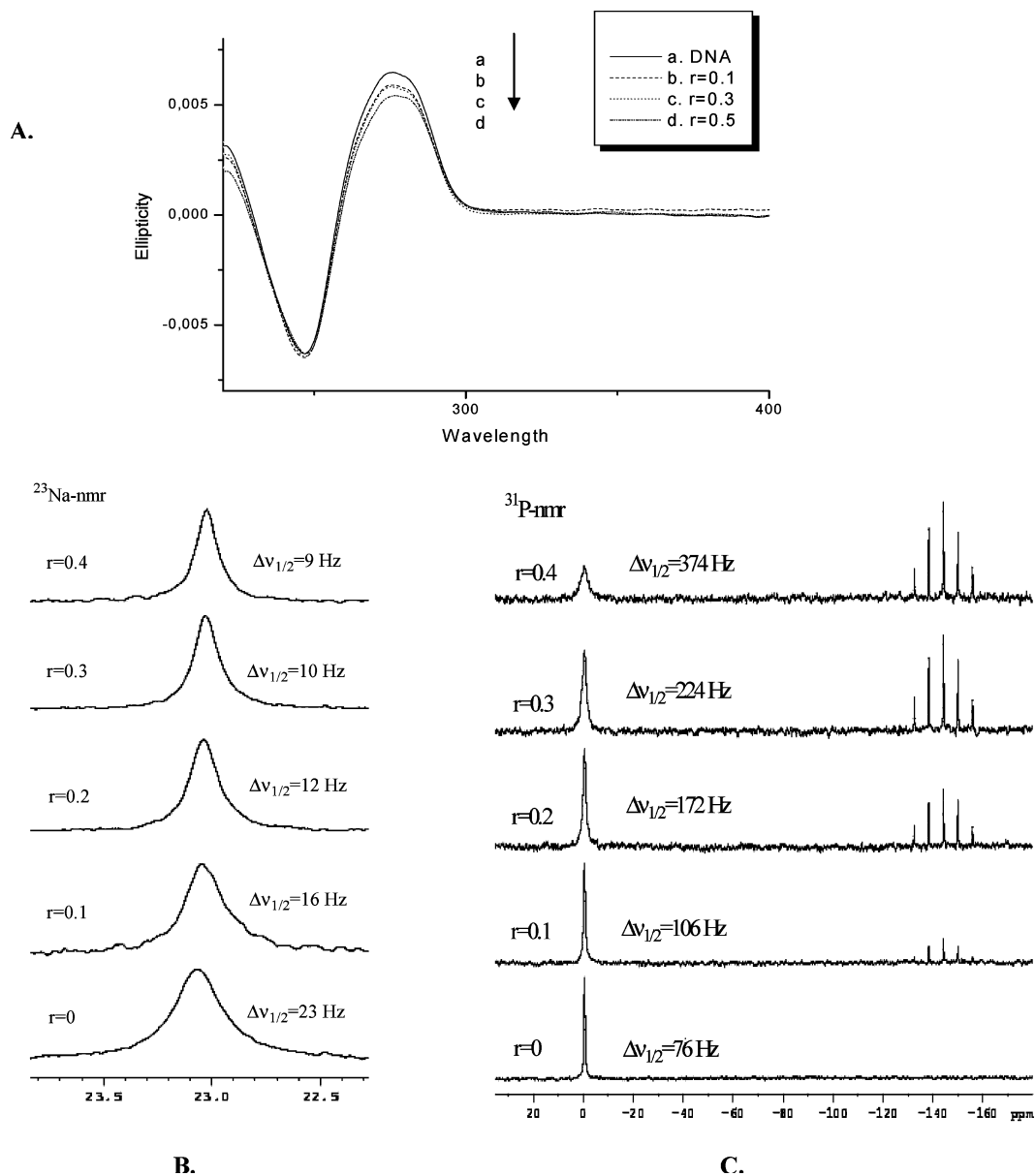


Figure 4. (A) Circular dichroism spectral changes of calf thymus DNA (70–300 bp) upon addition of complex **2** at increasing ratios. The arrow indicates the order from a to c. (B and C) ^{23}Na and ^{31}P NMR spectral changes corresponding to calf thymus DNA (70–300 bp) upon addition of complex **2** at increasing ratios.

arrangement of the $(\text{H}_2\text{O})_6$, the average $\text{O}\cdots\text{O}$ distance is 2.830 Å for complex **2**, which is very close to that observed by Ye⁴⁸ and slightly shorter than those observed from liquid water 2.854 Å.⁵¹ However, the value of the average angle $\text{O}\cdots\text{O}\cdots\text{O}$ is 117.60°, which is significantly different from the corresponding value of 109.3° in hexagonal ice.

The coordination polyhedron of Ru(II) is distorted octahedral and formed by five pyridinic nitrogen atoms (three nitrogen atoms from terpy and two from the ligand 4-CO₂-4'-Mebpy) and one more nitrogen atom from the anionic nitro group. The distortion from O_h symmetry is mainly expressed in the angles with cis angles ranging from 78.23(7)° to 101.87(8)° resulting in an angular variance⁵² of 59.50°. The $(\text{NO}_2)^-$ anion is coordinated through the

nitrogen, with a Ru–N bond length of 2.029(2) Å, which is among the shortest observed in the literature for similar complexes. For example, it is shorter than those observed for the complexes $[\text{Ru}^{\text{II}}(\text{trpy})(2,2'\text{-dipyridylamine})(\text{NO}_2)](\text{ClO}_4)$, 2.034 Å,⁵³ $[\text{Ru}^{\text{II}}(\text{trpy})(\text{arylazopyridine})(\text{NO}_2)](\text{ClO}_4)$, 2.057 Å,¹¹ $[\text{Ru}^{\text{II}}(\text{trpy})(\text{PMe}_3)_2(\text{NO}_2)](\text{ClO}_4)$, 2.074 Å,⁵⁴ and $[\text{Ru}^{\text{II}}(\text{trpy})(1\text{-Me-2-(2-pyridyl)1H-benzimidazole})(\text{NO}_2)](\text{ClO}_4)$.¹² The N(61)–O(62) and N(61)–O(63) bond distances are quite longer than those observed for similar complexes,^{12,55} probably due to the involvement of the O(63) in the hydrogen-bonding network. For the same reason, the C(27)–O(28) and C(27)–O(29) bond distances of the

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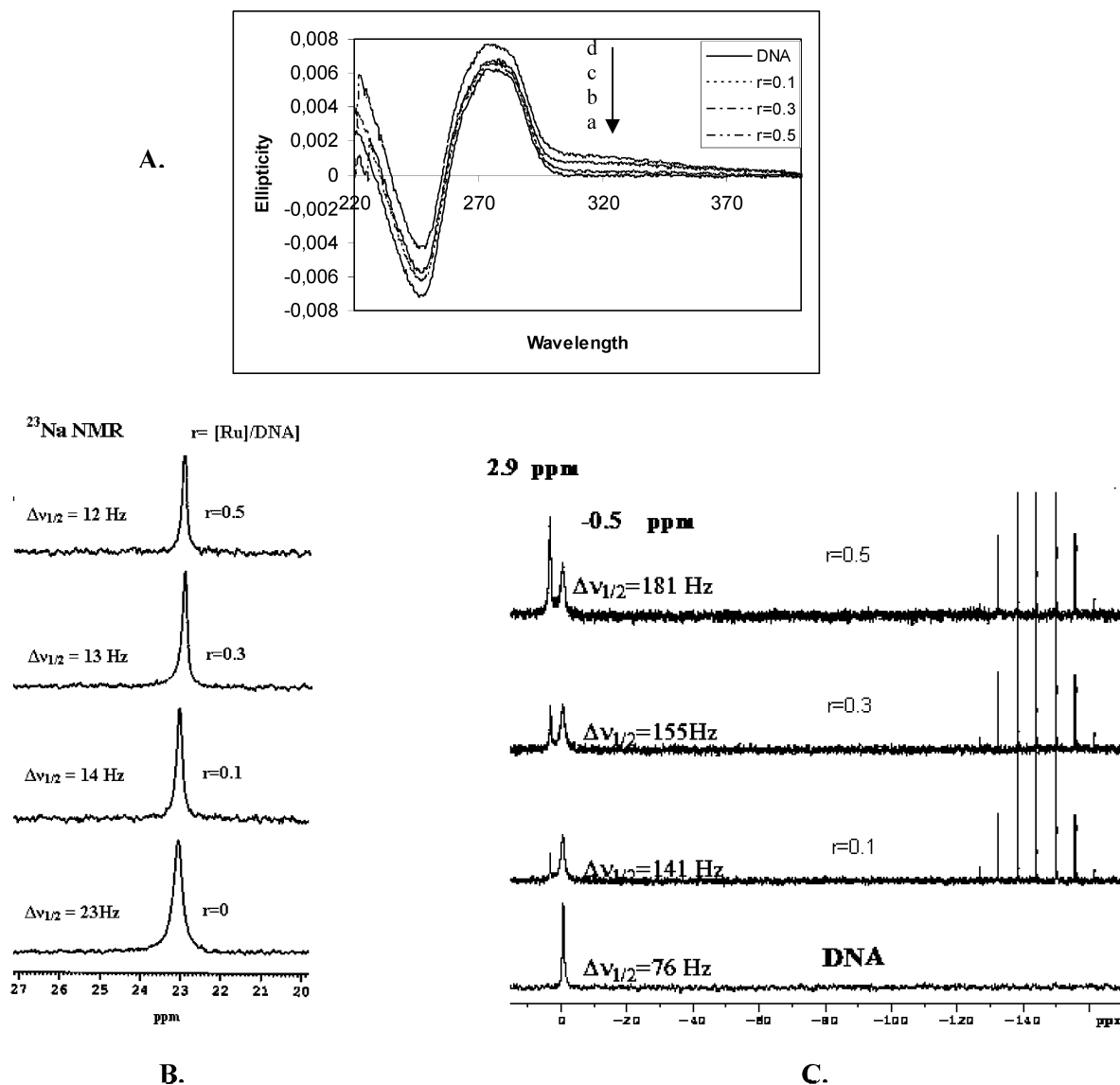


Figure 5. (A) Circular dichroism spectral changes of calf thymus DNA (70–300 bp) upon addition of complex **4** at increasing ratios. The arrow indicates the order from a to c. (B and C) ^{23}Na and ^{31}P NMR spectral changes corresponding to calf thymus DNA (70–300 bp) upon addition of complex **4** at increasing ratios.

carboxylic group are slightly longer than expected. The central pyridine ring of terpy is coordinated to metal center with a bond length of $\text{N}(41)\text{--Ru}(1) = 1.967(2) \text{ \AA}$ which is significantly shorter from the corresponding distances of the other two outmost terpy's pyridine rings. However, these values are in the average range for such complexes,^{11,53,54,12,55} and very similar to those observed for complex $[\text{Ru}^{\text{II}}(\text{trpy})\text{--}(4\text{-CO}_2\text{H-4}'\text{-Meppy})\text{Cl}]^+$.³⁵ As mentioned above (subsection iii), the ligand 4-CO₂H-4'-Meppy is able to coordinate to the metal center in two different orientations. The observation that in the isolated crystals the 4-CO₂H-4'-Meppy possesses a *trans*-(CO₂, NO₂) arrangement can be explained by the stability of dimers that are formed through hydrogen bonds which could not be formed in the case of the *cis* arrangement. In the case of complex $[\text{Ru}^{\text{II}}(\text{trpy})\text{--}(4\text{-CO}_2\text{H-4}'\text{-Meppy})\text{Cl}]^+$, where no dimers are formed even though the carboxylic group is protonated, the ligand 4-CO₂H-4'-Meppy prefers a *cis*-(Cl, CO₂) orientation.³⁵ The two Ru–N bond distances of the ligand 4-CO₂-4'-Meppy are not equal, with the Ru-

(1)–N(11) bond length being 2.096(2) Å and Ru(1)–N(21), 2.071(2) Å (Figure 3A). It has been suggested that such differences consist of a *trans* influence of the shorter Ru–N terpy bond (1.967(2) Å) to the opposite Ru–N bond, which is significantly elongated.³⁵ Similar behavior was also observed with complex $[\text{Ru}^{\text{II}}(\text{trpy})\text{--}(4\text{-CO}_2\text{H-4}'\text{-Meppy})\text{Cl}]^+$ where the Ru–N bond distance of the 4-CO₂H substituted pyridinic ring which is *trans* to the shorter Ru–N (terpy) bond is longer than the 4'-Me substituted ring.³⁵ However, in the *trans* position to the shorter Ru–N(terpy) pyridine ring of the ligand 4-CO₂-4'-Meppy is the 4'-Me-substituted and not the –CO₂H pyridine ring. This demonstrates clearly the minor role of the substitution in the pyridine ring, compared to the influence of the *trans*-effect.

(v) DNA Binding Studies. In order to investigate the roles of the conjugated peptide and the nitro group of the prepared complexes to their interactions with calf thymus DNA (70–300 bp), spectroscopic (CD, ^{23}Na and ^{31}P NMR) and analytical techniques were used. As mentioned above, the

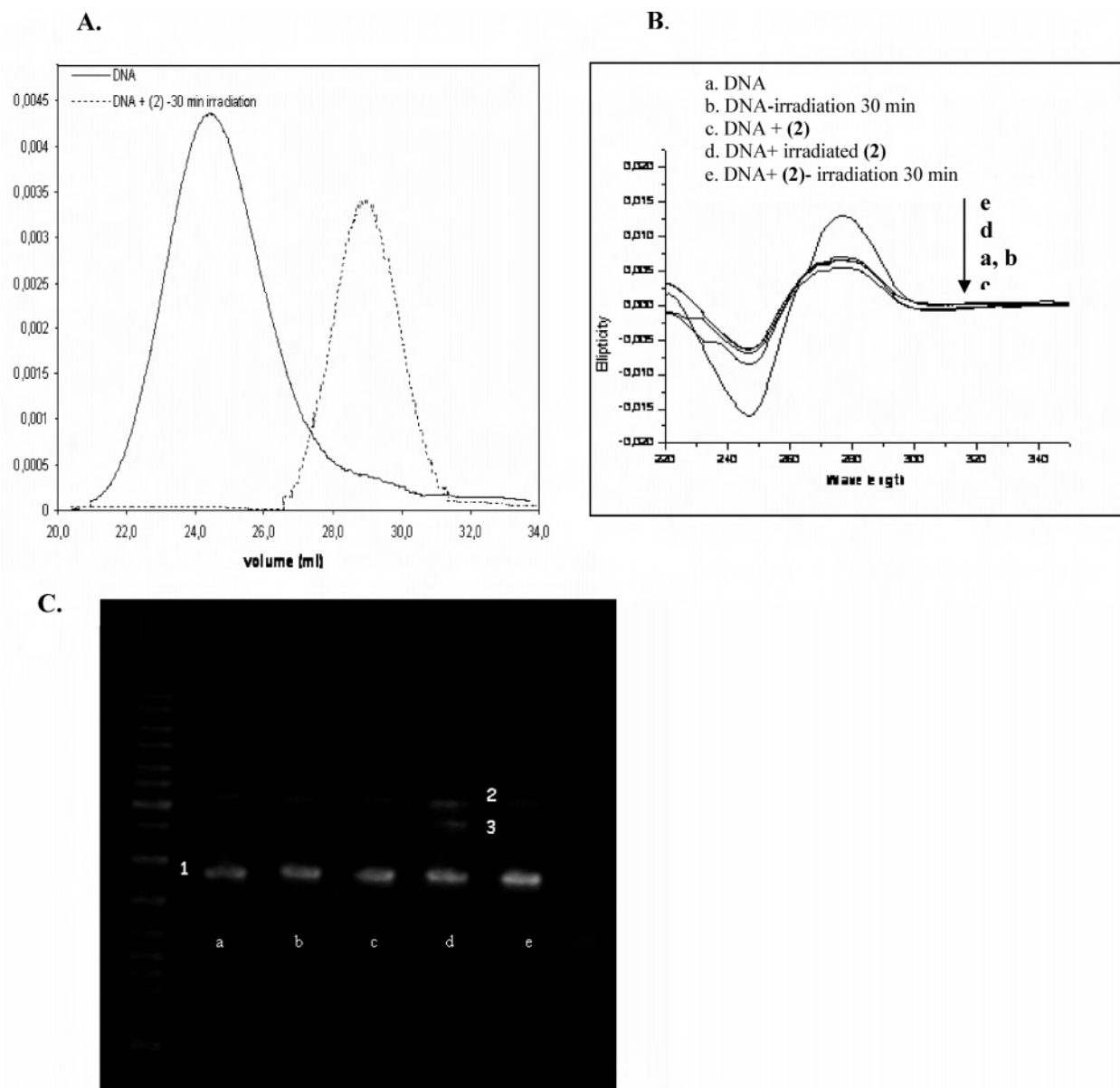


Figure 6. (A) Size exclusion chromatographic data of calf thymus DNA and the irradiated solution of DNA with complex **2**. (B) Circular dichroism spectral changes of calf thymus DNA (70–300 bp) upon addition of complex **2** and irradiation. The arrow indicates the order from a to e. (C). Electrophoretic mobility results of the plasmid DNA pUC9 upon addition of complex **2**: (a) supercoiled DNA; (b) irradiated supercoiled DNA for 30 min; (c) DNA–complex **2**, $r = 0.25$; (d) DNA–complex **2**, $r = 0.25$, irradiated for 30 min; (e) DNA-irradiated complex, lane 1, supercoiled DNA; lane 2, cleavage in single strand; lane 3, cleavage in double strand linear.

nitrosyl complexes **1** and **3** at neutral pH, where the DNA studies are performed, exist as nitro derivatives and for this reason only complexes **2** and **4** were studied.

The CD spectra of the short length calf thymus DNA, upon addition of the complexes at increasing ratios ($r = 0.1, 0.3$, and 0.5), are shown in Figure 4A. In general, complex **2** slightly affects the characteristic CD bands of B-type DNA, depending on the ratio. Upon ratio increase, the intensity of the positive band at 278 nm decreases (about 20% at ratio $r = 0.5$), while the band at 245 nm is remaining practically unaffected. Similar changes were obtained when double charged alkaline-earth metal cations interact at low concentrations with DNA, suggesting a shortening of the distance between the bottom of the major groove and the phosphate

groups.⁵⁶ Also, the triply positively charged cation $[\text{Co}^{\text{III}}(\text{NH}_3)_6]^{3+}$ has been found to induce similar changes in CD spectra of calf thymus DNA (160 and 3000–8000 bp) at low molar ratios (<0.3). In this case, a dehydration process of the DNA was suggested, and therefore a decrease of its volume.⁵⁷

The ^{23}Na NMR spectrum of complex **2** shows a line-width narrowing upon ratio increase from $r = 0$ to $r = 0.5$ ($\Delta\nu/2 = 23$ Hz to $\Delta\nu/2 = 8$ Hz) (Figure 4B). This effect is consistent with a perturbation of the Na^+ counterions of the DNA phosphate groups. The positively charged complex interacts with DNA, and a modification of the total charge

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of the DNA polyanion takes place. The replacing of the sodium ions from the DNA phosphates in complex **2** induces the narrowing in the ^{23}Na NMR signal. However, the ^{31}P NMR spectra under the same conditions (Figure 4C) show a progressive broadening of the phosphorus resonance and a slight (0.1 ppm) downfield shift, indicating an inflated dispersion of ^{31}P signals and a change in the ^{31}P environment. Taking into account that the DNA shortening observed by the circular dichroism data affects all the phosphate distances, it is suggested that the slight downfield shift may reflect the DNA shortening. The influence of the partial replacement of sodium counter ions, observed by the ^{23}Na NMR spectra, affects the surroundings of some phosphates, causing a dispersion of their chemical shifts. Also, it could be concluded that the reaction is in intermediately kinetics, in the NMR time scale at this temperature.

Complex **4** affects slightly both negative and positive CD bands of the DNA upon ratio increase (Figure 5A). The positive CD band at 278 nm is affected in a similar manner to complex **2**, suggesting a reduction of the DNA length. However, the intensity of the band at 245 nm decreases, indicating that conformational alterations induced in DNA by complex **4** are different from those induced by complex **2**. This observation was confirmed by the ^{31}P NMR spectra, which are different from those of complex **2**, where a new signal appears, 3.4 ppm downfield from the ^{31}P signal of the free DNA, accompanied by a broadening of the remaining signal (Figure 5C). Similar important downfield shifts were obtained in cases of strong DNA–ligand interactions, such as in the case of actinomycin,⁵⁸ where a new signal was observed, due to the slow kinetics of the reaction.⁵⁹ In the case of complex **4**, slow kinetics could arise from the formation of hydrogen bonds or other interactions between the peptides and DNA. Since the ^{23}Na NMR data of the DNA titration with complex **4** (Figure 5B) follow those of complex **2**, it is suggested that the peptide moiety binds to DNA in a synergistic way so that the whole complex binds.

(vi) Preliminary DNA Photocleavage Studies. Preliminary photocleavage studies of calf thymus and plasmid DNA were performed by irradiation of the DNA samples and complex **2** ($r = 0.1$) using a mercury lamp for 30 min. Size exclusion chromatography was used to analyze the photoproducts. Thus, no difference in the molecular weight of calf thymus DNA alone after 30 min of irradiation was observed (126 KD). However, the chromatograph of an irradiated aqueous solution of CT DNA and complex **2** under the same

conditions showed one peak corresponding to an average molecular weight of 30.2 KD, indicating that the DNA strands were photocleaved into shorter pieces (Figure 6A).

The CD spectrum of the irradiated calf thymus DNA and complex **2** shows a dramatic and almost equal increase in the intensities of both negative (245 nm) and positive (280 nm) DNA bands, without any other alteration in the spectrum. (Figure 6B). The pattern of the CD spectrum produced indicates clearly a B-type form for the nicked DNA.⁶⁰ In addition, no significant difference between the sample containing the separately irradiated **2** and DNA and the nonirradiated solution of DNA–complex **2** were observed (Figure 6B). The preliminary gel electrophoretic studies agree with the one of the CD spectra (Figure 6C).

A more detailed study of the photocleavage of CT DNA by complex **2** and other complexes is underway and will be published elsewhere.

Conclusions

In conclusion, this work has demonstrated the following: (i) The studied ruthenium nitrosyls are transformed to their nitro derivatives at neutral pH in solution. (ii) The photo-release of the NO in dry MeCN solutions produced solvolyzed photoproducts of Ru(III) which were detected. (iii) Complex **2** interacts with DNA and shortens slightly the DNA length. (iv) The conjugation of the peptide GHK to complex **2**, however, results in a relatively tighter binding to DNA, with the peptide moiety acting in a synergistic way with the whole complex. Preliminary results have indicated photocleavage of CT DNA.

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